

# VitroView<sup>TM</sup> Anti-Goat Immunofluorescenc Staining Kit (Green, 50-100 slides)

SKU#: VB-6109

# **Description:**

Immunofluorescence (IF) is a vital immunochemical technique enabling the detection and precise localization of numerous antigens across diverse tissue types and various cell preparations. This VitroView<sup>TM</sup> Anti-Goat Immunofluorescence Staining Kit is a reliable and user-friendly solution designed for use with primary antibodies specifically raised in goat.

The advantages of this kit include: 1) High sensitivity; 2) Low background; 3) Reduction of steps and time; 4) Simplified multiple labeling.

# **Specifications**

Unit Size	1 kit
Target Species	Goat
Conjugates and Color of	FL488 conjugated donkey anti-goat IgG
Fluorescence	(Green)
Excitation/Emission for FL488 dye	496/519 nm
<b>Host Species</b>	Donkey
Application	Immunofluorescence staining for detecting a
	primary antibody made in goat

#### **Application**

Immunofluorescence staining for detecting a primary antibody made in goats

### **Kit Contents:**

- RTU Normal Donkey Serum -----10ml
- FL488 Conjugated Donkey Anti-Goat IgG (1mg/ml)-----50µl
- Aqueous Anti-fade Mounting Medium with Dapi-----1.5 ml×2

#### Storage

Store at 2-8 °C.

## Reagents and Material Required but Not Provided

- Xylene and ethanol for FFPE samples
- Distilled or deionized water
- 10 mM phosphate-buffered saline (PBS), pH 7.4
- Triton X-100
- Mini PAP Pen

- Primary antibody
- Antibody Dilution Buffer (SKU#: VB-6002)
- BSA
- Antigen retrieval reagents

#### **Protocol**

# 1. Preparation of Slides

#### A. Cell Lines

- Grow cultured cells on chamber slides, sterile glass cover slips or slides overnight at 37°C.
- Briefly wash with PBS.
- Choose one of the following fixation methods (To achieve optimal results, it may be necessary to optimize the fixation methods).
  - a. Fix in 10% formalin in PBS for 20 minutes (keep wet).
  - b. Immerse in ice-cold methanol for 10 minutes and allow to air dry.
  - c. Immerse in ice-cold acetone for 10 minutes and allow to air dry.
- Wash slides in PBS.

#### **B. Frozen Sections**

- Snap-freeze fresh tissues in liquid nitrogen or isopentane pre-cooled in liquid nitrogen, then embed them in OCT compound in cryomolds. Store the frozen tissue block at -80°C until ready for sectioning.
- Before sectioning, equilibrate the frozen tissue block to the temperature of the cryotome cryostat (e.g., -20°C).
- Section the frozen tissue block into the desired thickness (typically 5-10 μm) using the cryotome.
- Place the tissue sections onto glass slides suitable for immunohistochemistry (e.g., Superfrost).
- Store sections in a sealed slide box at -80°C for later use.
- Before staining, allow slides to warm to room temperature for 30 minutes and then fix in ice-cold acetone or ice-cold methanol for 10 minutes. Air dry for 30 minutes.
- Wash in PBS.

## C. Paraffin Sections

- Deparaffinize sections in xylene, 3×5 minutes.
- Hydrate with 100% ethanol, 2×2 minutes.
- Hydrate with 95% ethanol, 2×2 minutes.
- Rinse in distilled water.
- Follow the required pretreatment procedure.

# 2. Antigen Retrieval

Web: <u>www.vitrovivo.com</u>. Phone: 301-500-0499 Email: orders@vitrovivo.com Most formalin-fixed tissues require antigen retrieval before proceeding with immunohistochemical staining. Common methods include heat-mediated and enzymatic antigen retrievals. Choose the appropriate method:

- a. For Citrate: Bring slides to a boil in 10 mM sodium citrate buffer, pH 6.0, and maintain at a sub-boiling temperature for 10 minutes. Cool slides on the benchtop for 30 minutes.
- b. For EDTA: Bring slides to a boil in 1 mM EDTA, pH 8.0, and follow with 15 minutes at a sub-boiling temperature. No cooling is necessary.
- c. For TE: Bring slides to a boil in 10 mM TE/1 mM EDTA, pH 9.0, and then maintain at a sub-boiling temperature for 18 minutes. Cool at room temperature for 30 minutes.
- d. For Pepsin: Digest for 10 minutes at 37°C.

Note: Do not use this pretreatment with frozen sections or cultured cells that are not paraffinembedded.

## 3. Staining Procedure

- a. Rinse sections in PBS-Triton X-100 (0.025%) for two 2-minute cycles.
- b. Serum Blocking: Incubate sections with 2-4 drops of ready-to-use (RTU) normal donkey serum for 30 minutes to block non-specific immunoglobulin binding.
- c. Primary Antibody: Incubate sections with primary goat antibody at the appropriate dilution in antibody dilution buffer (SKU# #: VB-6002) for 1-2 hours at room temperature or overnight at 2-8°C. Rinse in PBS.
- d. Detection: Incubate sections with FL488-conjugated Donkey Anti-Goat IgG secondary antibody at suitable dilutions (ranging from 1/100 to 1/500) in 0.1% BSA in PBS for 1 hour at room temperature while keeping the samples in the dark.
- e. Rinse in PBS for three 2-minute cycles.
- f. Counterstaining and mount cover slip with 20-30µl of aqueous anti-fade mounting medium with DAPI. Seal cover slip with nail polish to prevent drying and movement.
- g. Visualize and capture the signal using a fluorescence microscope equipped with excitation wavelength filters of 488 nm.
- h. Store slides in the dark at 2-8°C.

Note: This product is intended for research purposes only. This product is <u>not</u> intended to be used for therapeutic or diagnostic purposes in humans or animals.

Precautions: Handle with care. Avoid contact with eyes, skin and clothing. Do not ingest. Wear gloves.

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