



**VitroView™ DAPI Nuclear Stain Reagent**  
**SKU#: VB-1000**

**Description:** The nucleus is a membrane-bound organelle found in eukaryotic cells. Cell nuclei contain most of the cell's genetic material (chromosome), organized as multiple long linear DNA molecules complexed with various proteins, such as histones, to form chromosomes. The nucleus is the primary site of gene expression and DNA replication, within which is a sub-compartment known as the nucleolus responsible for synthesizing rRNA and assembling ribosomes.

4',6-diamidino-2-phenylindole (DAPI) is a popular fluorescent dye for nuclear and chromosome counterstain. It emits blue fluorescence (Ex:358nm/Em:461nm) upon binding to AT rich regions in DNA. At higher concentrations DAPI can pass through an intact cell membrane, therefore it can be used to stain both live and fixed cells and provides a marker for membrane viability.

**Contents:**

VB-1000 RTU DAPI Nuclear Staining Solution-----30 ml

**Storage**

Store at 2-8°C and protect from light.

**Material Needed But NOT Supplied with the Kit:**

1. Staining jar
2. Glass slides
3. PBS (Wash Buffer)
4. Mount media

**General Protocol:**

Adherent cells for fluorescence microscopy

1. Grow cultured cells on sterile glass cover slips or chamber slides overnight at 37 °C.
2. Follow appropriate protocol to fix cultured cells.
3. Completely wash the cells with PBS as needed.
4. Add adequate RTU DAPI Nuclear Staining Solution to cover the whole sample.
5. Incubate under dark at room temperature for 1–5 minutes.
6. Rinse the sample several times with PBS and remove excess DAPI.
7. Add antifade aqueous mounting medium and mount.
8. Use appropriate filters (Ex:358nm/Em:461nm) and detect under fluorescence microscope according to standard protocol.

Suspension cells for fluorescence microscopy

1. The cells are harvested into a 15 mL polypropylene centrifuge tube and spin down for 8 min at 600 RPM,
2. The supernatant is discarded and the cells are resuspended in 0.5 ml of culture medium

3. 1-2 drops of the cell suspension were placed on a slide in the central area and moved around to form a thin and even film with a glass spreader.
4. (Option) You also can use cytocentrifuge to prepare cell slides.
5. Air dry and follow appropriate protocol to fix cultured cells.
6. Drop adequate RTU DAPI Nuclear Staining Solution to cover the whole sample on slide.
7. Incubate under dark at room temperature for 1–5 minutes.
8. Cover with coverslip and view under fluorescence microscope according to standard protocol.

For paraffin sections

1. Deparaffinize sections in xylene, 3×5min.
2. Hydrate with 100% ethanol, 2×2min.
3. Hydrate with 95% ethanol, 2×2min.
4. Rinse in distilled water.
5. Add adequate RTU DAPI Nuclear Staining Solution to cover the whole sample.
6. Incubate under dark at room temperature for 1–5 minutes.
7. Rinse the sample several times with PBS and remove excess DAPI.
8. Add antifade aqueous mounting medium and mount.
9. Use appropriate filters and detect under fluorescence microscope according to standard protocol.

For frozen sections

1. Before staining, warm frozen slides at room temperature for 10-30 minutes.
2. Fix in ice cold acetone or ice cold methanol for 10 minutes.
3. Air dry for 30 minutes and wash in PBS.
4. Add adequate RTU DAPI Nuclear Staining Solution to cover the whole sample.
5. Incubate under dark at room temperature for 1–5 minutes.
6. Rinse the sample several times with PBS and remove excess DAPI.
7. Add antifade aqueous mounting medium and mount.
8. Use appropriate filters and detect under fluorescence microscope according to standard protocol.

**Note:** This product is intended for research purposes only. This product is not intended to be used for therapeutic or diagnostic purposes in humans or animals.