



VitroView™ *In Situ* LC3B Autophagy Detection Kit
(For 50 Tests)
SKU#: VB-4004D

Description

Autophagy (or autophagocytosis) is the basic catabolic mechanism that involves cell degradation of unnecessary or dysfunctional cellular components through the actions of lysosomes. Transmission electron microscopies (TEM) as well as immunohistochemistry are indispensable tools for the evaluation of autophagy *in situ*. LC3B is currently considered as one of the most reliable markers of the autophagic process. The *in situ* LC3B autophagy detection kit has been optimized for the detection of the autophagy marker protein LC3B in paraffin-embedded tissue specimens, cultured cells and frozen sections. The antibody against LC3B works in human, mouse, and rat with strong specific staining and lower background.

Application

In situ detection of autophagy cells in human, mouse, and rat cells/tissues

Contents

1. 10×Ag retrieval solution-----50 ml
2. RTU Block buffer-----5 ml
3. RTU anti-LC3B antibody -----5 ml
4. RTU biotinylated anti-rabbit secondary antibody -----5 ml
5. RTU streptavidin-HRP-----5 ml
6. DAB Stock Solution (40×)-----0.5 ml
7. Stable H₂O₂ Solution (40×)-----0.5 ml
8. DAB Buffer-----15 ml
9. LC3B positive control FFPE slides-----2 slides

Note: RTU=ready-to-use

Reagents and Material Required but Not Provided

- Xylene and ethanol
- Distilled or deionized water
- 30% hydrogen peroxide
- 10 mM phosphate-buffered saline (PBS), pH 7.4
- Triton X-100
- Mini PAP Pen
- Hematoxylin (Cat#:VB-6004)
- Mounting Media

Storage

Store at 2-8°C.

Protocol:

1. Preparation of Slides

1). Cell Lines

- a. Grow cultured cells on sterile glass cover slips or slides overnight at 37 ° C
- b. Wash briefly with PBS
- c. Fix as desired. Possible procedures include:

- a. 20 minutes with 10% formalin in PBS (keep wet)
- b. 10 minutes with ice cold methanol, allow to air dry
- c. 10 minutes with ice cold acetone, allow to air dry
- d. Wash in PBS

2). Frozen Sections

- a. Snap frozen fresh tissues in liquid nitrogen or isopentane pre-cooled in liquid nitrogen, embedded in OCT compound in cryomolds. Store the frozen tissue block at -80°C until ready for sectioning.
- b. Transfer the frozen tissue block to a cryotome cryostat (e.g. -20°C) prior to sectioning and allow the temperature of the frozen tissue block to equilibrate to the temperature of the cryotome cryostat.
- c. Section the frozen tissue block into a desired thickness (typically 5-10 µm) using a cryostat.
- d. Place the tissue sections onto glass slides suitable for immunohistochemistry (e.g. Superfrost).
- e. Sections can be stored in a sealed slide box at -80°C for later use.
- f. Before staining, warm slides at room temperature for 30 minutes and fix in ice cold acetone or ice cold methanol for 10 minutes. Air dry for 30 minutes.
- g. Wash in PBS

3). Paraffin Sections

- a. Deparaffinize sections in xylene, 3×5min.
- b. Hydrate with 100% ethanol, 2×2min.
- c. Hydrate with 95% ethanol, 2×2min.
- d. Rinse in distilled water.
- e. Follow procedure for pretreatment as required.

2. Antigen retrieval

Formalin-fixed tissues require an antigen retrieval step before immunohistochemical staining can proceed.

- 1) Dilute 10×Ag Retrieval Solution to 1×Ag Retrieval Solution by adding 90ml ddH₂O into 10ml of 10×Ag Retrieval Solution.
- 2) Bring slides to a boil in 1×Ag retrieval solution at a sub-boiling temperature for 10-15 minutes. Cool slides on bench top for 30 minutes.

Note: Do not use this pretreatment with frozen sections or cultured cells that are not paraffin-embedded.

3. Staining Procedure

- 1) Rinse sections in PBS-Triton X-100 (0.025%) for 2×2min
- 2) **Serum Blocking:** incubate sections with 2-3 drops of RTU normal goat serum for 30 minutes to block non-specific binding of immunoglobulin.
- 3) **Primary Antibody:** incubate sections with RTU anti-LC3B primary antibody at room temperature for 1-2 hour or at 4 °C for overnight. Rinse in PBS.
- 4) **Peroxidase Blocking (optional):** incubate sections in 0.3% hydrogen peroxide in PBS for 10 minutes at room temperature. Rinse in PBS.
- 5) **Secondary Antibody:** incubate sections with 2-3 drops of RTU biotinylated anti-rabbit secondary antibody for 30 minutes at room temperature.
- 6) Rinse in PBS for 3×2min.
- 7) **Detection:** incubate sections with 2-3 drops of RTU streptavidin-HRP for 30 minutes at room temperature.
- 8) Rinse in PBS for 3×2min.
- 9) **Chromogen/Substrate:** incubate sections with 3 drops of DAB solution for 2-8 minutes. Monitor signal development under a microscope
Note: DAB solution is made by mixture of 20 µl of DAB stock solution and 20 µl of stable H₂O₂ solution with 1ml of DAB buffer (brown stain). Rinse in distilled water 2×2 min
- 10) **Counterstain:** For using Hematoxylin Nuclear Counterstaining Kit (CAT#: VB-6004), incubate sections with 3 drops of RTU hematoxylin solution for 1-2 minutes. Rinse in tape water 2×2 min.
- 11) Dehydrate through 75% ethanol for 2 min, 95% ethanol for 2 min, and 100% ethanol for 2×3min. Clear in xylene for 2×5min.
- 12) Coverslip with mounting medium.

IHC Troubleshooting

High background staining

| Possible Cause | Solution |
|----------------------------------------------------------|------------------------------------------------------------------------------------------------------------------------------------------------|
| Endogenous peroxidase activity was incompletely blocked. | Incubate sections in 0.3% hydrogen peroxide in methanol or PBS for 10-30 minutes at room temperature. |
| Deparaffinization was incomplete. | Prepare new sections and deparaffinize according to standard laboratory protocol using fresh xylene or xylene substitute. |
| Inadequate rinsing of slides. | Gently rinse slide with wash buffer bottle and place in wash bath for 5 minutes. Gentle agitation of the wash bath may increase effectiveness. |
| Over-development of substrate. | Reduce incubation time. |
| Dehydration of specimen during staining. | Keep section wet. |

Negative staining on positive slides

| Possible Cause | Solution |
|----------------------------------------------------------------------|-----------------------------------------------------------------------------------------------------------------------------------------|
| Steps in the staining protocol were performed in incorrect sequence. | Repeat the procedure. |
| Primary or secondary antibody incubation steps were omitted. | Repeat the procedure. |
| Labile antigens were destroyed. | Use fresh cutting slides. Use a paraffin wax with a melting temperature ~55-58°C. Wax used for embedding should not be exceed 60 °C. |
| Specimen was improperly fixed and/or processed. | Check manufacture's specifications regarding recommended fixative |
| Specimen dehydrated during staining. | Repeat the procedure by following the manufacture's protocol. |

Weak staining on all slides

| Possible Cause | Solution |
|------------------------------------------------------------------------------------|---------------------------------------------------------------------------------------------------------------------------|
| Specimen retained excess liquid after rinsing steps. | Remove excess liquid after rinsing steps. |
| Incubation times were insufficient. | Prolong incubation time. |
| Substrate prepared improperly. | Check compatibility of buffer ingredients with enzyme and substrate-chromogen reagents. Repeat staining. |
| Deparaffinization was incomplete (staining may be accompanied by high background). | Prepare new sections and deparaffinize according to standard laboratory protocol using fresh xylene or xylene substitute. |

Warning: DAB is a possible carcinogen. Please take necessary precautions.

Note: This product is intended for research purposes only. This product is not intended to be used for therapeutic or diagnostic purposes in humans or animals. Avoid contact with eyes, skin and clothing. Do not ingest. Wear gloves.